Changes in α -tocopherol and retinol levels during cardiopulmonary bypass correlate with maximal arterial partial pressure of oxygen

IRENE VALLE-GINER¹, EZEQUIEL MARTÍ-BONMATÍ², AMPARO ALEGRÍA-TORÁN³, ANASTASIO MONTERO⁴, & ESTEBAN J. MORCILLO⁵

¹Pharmacy Service, 'Obispo Polanco' General Hospital, Teruel, Spain, ²Pharmacy Service, General University Hospital Consortium, Valencia, Spain, ³Department of Nutrition and Food Chemistry, Faculty of Pharmacy, University of Valencia, Valencia, Spain, ⁴Cardiac Surgery Services, General University Hospital Consortium, Valencia, Spain, and ⁵Clinical Pharmacology Unit and Department of Pharmacology, University Clinic Hospital, Faculty of Medicine, University of Valencia, Valencia, Spain

Accepted by Professor G. Mann

(Received 25 January 2007; in revised form 30 April 2007)

Abstract

Cardiopulmonary bypass (CPB) is associated with oxidative stress. This study examined antioxidant levels in adults undergoing CPB surgery and their correlation with clinical variables. Arterial blood samples were obtained from 27 patients undergoing CPB. The time-course variation of vitamin C (spectrofluorimetry), α -tocopherol and retinol (HPLC) levels were determined. Plasma vitamin C rose initially but gradually decayed during reperfusion until 60% reduction of baseline values post-surgery. α -Tocopherol and retinol were reduced along CPB with post-operative values $\sim 25\%$ lower than baseline. No significant changes were found for selenium and glutathione peroxidase. PaO₂ values rose steadily throughout CPB. A correlation existed for α -tocopherol and retinol depletion vs maximal PaO₂ throughout CPB but no correlation was found for antioxidant consumption vs duration of ischaemia and reperfusion and hypothermia level. In conclusion, consumption of arterial blood antioxidant vitamins occurs with CPB in relation with PaO₂ levels but not for other clinical variables measured in this study.

Keywords: Cardiopulmonary bypass, oxidative stress, antioxidants, vitamin C, α -tocopherol, retinol

Introduction

An increasing number of patients are undergoing cardiac surgery and in most cases cardiopulmonary bypass (CPB) is required [1]. This procedure involves diverting blood from the heart and then returning it after oxygenation to a high-capacity artery, thus creating a bypass which functions in series with the circulation to the rest of the body. CPB is an aggressive technique frequently associated with post-surgical complications including myocardial, pulmonary, renal, liver and neurological dysfunction, coagulation disorders and, ultimately, multiple organ failure [2] in which perioperative oxidative stress appears implicated [3–6]. Reactive oxygen species originate from various cellular and enzymatic sources such as myocardial cells, activated neutrophils and endothelial xanthine oxidase [3]. Many studies have described the nature of these oxidant species and the time course of their formation during CPB [3–7]. The oxidative stress is usually the result of an increased formation of reactive oxygen species which is not appropriately balanced by the wide spectrum of endogenous antioxidant defences [8]. Thus, in order

Correspondence: Dr Ezequiel Martí-Bonmatí, Pharmacy Service, University General Hospital Consortium, Av. Tres Cruces s/n, E-46014 Valencia. Spain. Tel: 34-961972000x72141. Fax: 34-963798306. E-mail: marti_eze@gva.es

to counterbalance this sequence of events and to diminish oxidative damage, several studies have investigated the use of exogenous antioxidant supplements during CPB [9–11]. However, although the systemic increase in oxidative stress during CPB and ischemia-reperfusion is well-documented [4–6, 12-14], the relative contribution of the various components of the endogenous antioxidant system and their correlation with clinical variables remains unsettled and continues being the object of further investigation.

In the present study, we have selected ascorbic acid, α -tocopherol, retinol, selenium and glutathione peroxidase activity as relevant markers of endogenous antioxidant defences. These markers were examined in arterial blood samples obtained at different times during adult CPB to study the temporal sequence of oxidative events that occur during this surgical procedure and if any correlation exists with some clinical variables.

Materials and methods

Patients and study design

The study protocol conforms the principles outlined in the Declaration of Helsinki (Edinburgh, 2000) and was approved by the Ethical Committee of the General University Hospital Consortium (Valencia, Spain) and written informed consent was obtained prior to enrolment. A total of 27 patients with elective, first-time cardiac surgery with CPB were included in this study. Cardiovascular and lipidlowering medication taken by these patients was the conventional therapy for their associated pathologies and not reported to affect antioxidant levels. The patients enrolled in this study showed no indication of impaired nutritional status according to physical examination, blood cells and chemistry and absence of significant previous weight loss. All patients reported standard dietary habits. Patients had received no dietary supplements including vitamins or antioxidant drugs and had no evidence of a systemic inflammatory response (C-reactive protein $\leq 10 \text{ mg/l}$ [15]. Patients receiving blood transfusions before or during surgery were excluded. Patients were given antibiotic prophylaxis before surgery.

After standard intravenous anaesthesia, CPB was established by cannulation of the ascending aorta and right atrium, with moderate hypothermia and haemodilution as previously outlined [16]. Myocardium was preserved with cold anterograde-retrograde crystalloid multidose cardioplegia using a standard potassium-based solution (Cardibraun[®], Braun, Melsugen, Germany; osmolarity 657 mOsm/l) mixed with blood at a ratio of 1:4. Surgery was performed using the intermittent cross-clamp technique. After decannulation, heparin was neutralized with protamine chloride (1:1 ratio).

Blood samples were collected from the radial artery or from the arterial side of pump during the extracorporeal circulation as follows: (i) baseline, immediately after cannulation of the arterial system before cardiac surgery (T_0) ; (ii) 3 min after initiating CPB but before a rtic clamp (T_1) ; (iii) during the ischemic period of CPB, at 3 and 30 min after aortic crossclamping (T_2, T_3) ; (iii) during reperfusion, with samples obtained 10 min before (T_4) and 15 min after (T_5) switching pump off; and (iv) 3 h after cessation of CPB with the patient within the intensive care unit (T_6) . Blood was drawn into plastic syringes, immediately transferred to tubes with or without EDTA (Vacutainer, Becton Dickinson, Inc., Oxnard, CA) as appropriate, placed on ice and protected from daylight. Samples with hemolysis were excluded from analysis. Blood samples were processed within 10 min after sampling to avoid auto-oxidation of endogenous antioxidants [3]. After centrifugation at 3000 g for 10 min at 4° C, the supernatant was distributed into aliquots and tubes were immediately frozen and stored at -20° C until analysis. Measurements were performed within 4 h after blood sampling for vitamin C, 48 h for selenium and within 14 days for the rest of the analytes.

In separate experiments, we demonstrated that analyte concentrations remained stable during this time of storage (not shown). A separate aliquot of EDTA-blood was centrifuged in microcapillaries (10000 g for 3 min, 22°C) for the determination of the hematocrit (HCT, ratio of erythrocytes to total blood volume). Correction to account for the haemodilution secondary to the pump-priming solution used for the extracorporeal CPB circuit was necessary. Thus, the concentrations of analytes at a particular time point were multiplied by the ratio of the initial hematocrit to the hematocrit at the specified time.

Analytical methods

Plasma vitamin C (ascorbate and dehydroascorbate) was analysed in *meta*-phosphoric acid-stabilized plasma by spectrofluorimetry [17]. Serum retinol and α -tocopherol were quantified by ultraviolet detection at 292 nm after extraction into hexane and separation by reversed-phase high performance liquid chromatography [18]. Plasma selenium was determined by flameless atomic absorption spectro-photometry (Perkin Elmer series 4100ZL, with Zeeman background correction and stabilized temperature platform furnace STPF) [19]. Plasma gluta-thione peroxidase (GPx) activity was estimated using a spectrophotometric assay (340 nm) according to the FLAIR protocol [20].

In additional experiments we assured that (i) concentrations of the different analytes in the samples were above the limit of detection of the corresponding

techniques, (ii) intra- and inter-assay coefficients of variation of the assays were below 5 and 10%, respectively, and (iii) determinations repeated at intervals along a 12 h period of time after samples were unfrozen yielded reproducible values with variation below 5% (not shown).

Statistical analysis

Values are expressed as mean \pm SD. Normality of variable distribution was assessed by the Shapiro-Wilk test. One-way ANOVA was used to test the time-dependent change in oxidative stress markers. *Post-hoc* comparisons were performed with Bonferroni adjustment for multiple comparisons. Correlations between variables were assessed by linear regression analysis. Statistical analysis was carried out using the software SPSS 9.0 for Windows (SPSS Inc., IL). p < 0.05 was considered significant.

Results

Patient characteristics

The main clinical characteristics of patients and a summary of their operative data are shown in Table I.

Table I. Clinical and operative data of 27 adult patients undergoing cardiac surgery with cardiopulmonary bypass (CPB).

Clinical data	
Age (years; mean \pm SD)	64.7 ± 9.36
Sex, male:female (%)	19 (70.4):8 (29.6)
Body weight (kg)	77.2 ± 14.9
Height (cm)	164 ± 10
Body surface (m ²)	1.83 ± 0.23
Smoking status,	14 (51.9):4 (14.8):
no:yes:ex-smoker:lost (%)	8 (29.6):1 (3.7)
Alcohol consumption, no:yes:lost (%)	25 (92.6):1 (3.7):1 (3.7)
NYHA class ^a , II:III:IV:lost (%)	2 (7.4):19 (70.4):
	1 (3.7):5 (18.5)
ASA class ^b	21 (77.8):1 (3.7):5 (18.5)
Associated pathologies ^{c,d}	4:5:9:5:3:4
Operative data	
Surgical procedure ^e , CABG:VR	20:7
CPB time (min)	116 ± 36
Aortic cross-clamp time (min)	77 ± 26
Reperfusion time (min)	39 ± 19
Temperature during hypothermia	29.8 ± 1.9
Maximal PaO ₂ (mmHg) ^f	387 ± 76
Administered volume ^g (ml)	3341 ± 474

^{*a*} New York Heart Association; ^{*b*} American Society of Anesthesiology; ^{*c*} insulin-dependent diabetes:non-insulin-dependent diabetes: hypercholesterolemia:hypertension:chronic obstructive pulmonary disease; ^{*d*} the total number of cases is greater than 27 because some patients have more than one associated pathology; ^{*c*} coronary artery bypass grafting:valve replacement; ^{*f*} Maximal PaO₂ values significantly increased after oxygenation from baseline values (T_0) of 85.4±12 up to 280±63 (T_1), 325±60 (T_2), 370±72 (T_3), 387±76 (T_4) and 373±78 (T_5) and decreased post-surgery to 122±35 (T_6); ^{*g*} cardioplegic solution and priming of the pump.

CPB is associated with changes in endogenous antioxidant levels

Pre-operative levels of vitamin C in the population studied were mostly normal except for 3 patients (11%) with marginal deficiency status (values close to 20 μ M) [4,21]. Vitamin C levels tended to increase during the initial period of CPB and ischemia followed by a gradual decrease during reperfusion (Figure 1A). Post-surgery haemodilution-corrected ascorbate levels were ~ 60% lower than baseline levels.

All patients in our study showed serum α -tocopherol and retinol pre-operative values within normal limits [22]. Serum α -tocopherol was reduced from early stages of CPB and during ischemia and reperfusion with the lowest valued reached after surgery (Figure 1B). Post-surgery haemodilution-corrected α -tocopherol levels were $\sim 24\%$ lower than baseline levels. Similar changes were observed for serum retinol with $\sim 27\%$ post-surgical decrease compared to pre-operative values (Figure 1B).

Plasma selenium and GPx activity did not significantly change during and after CPB (Figure 2).

Correlations between changes in plasma levels of antioxidants

The observed changes in plasma vitamin C concentrations were correlated with the corresponding changes in plasma α -tocopherol (p = 0.043) but the r value obtained (r = 0.146) indicates a weak correlation (Figure 3A). Correlation between vitamin C and retinol did not reach statistical significance (p = 0.058, r = 0.138) (Figure 3B). A significant correlation was found between plasma changes of α -tocopherol and retinol (p = 0.0001, r = 0.566) (Figure 3C).

There was a direct correlation between preoperative vitamin C concentrations and the extent of its decrease from baseline in absolute values after CPB (p = 0.002, r = 0.633); i.e. the higher the pre-operative value, the greater the corresponding post-operative fall in plasma vitamin C. By contrast, no significant correlation was found between preoperative values and their corresponding amounts of loss at post-operative time points for α -tocopherol (p = 0.325, r = 0.196) and retinol (p = 0.669, r =0.092).

Correlations between post-operative loss in plasma antioxidants and clinical parameters

The amount of loss of vitamin C after cardiac surgery did not correlate with CPB time (p = 0.372, r = 0.189), ischemic time (p = 0.257, r = 0.236), reperfusion time (p = 0.896, r = 0.030), hypothermia (p = 0.553, r = 0.128) or PaO₂max (p = 0.125, r = 0.315; Figure 4A). The amount of loss of α -tocopherol and retinol was significantly correlated with PaO₂max



Figure 1. Time-course of changes in concentration of vitamin C (A) and α -tocopherol and retinol (B) in arterial blood from adult patients subjected to cardiac surgery with bypass (CPB). Samples were obtained pre-operatively (T_0), after commencing the CPB (T_1), during ischemia (T_2 , T_3) and reperfusion (T_4 , T_5) and after surgery (T_6), as indicated in Methods. Data are mean \pm SD of 27 patients. *p < 0.05 from baseline values.

(p=0.047, r=0.395 and p=0.014, r=0.467, respectively) but did not correlate with duration of CPB, ischemia and reperfusion nor with hypothermia (Figure 4B and C).

Discussion

In the present study, we examined the temporal profile of arterial blood antioxidant vitamins in a cohort of 27 adult patients undergoing elective heart surgery with CPB and compared these changes to some clinical variables. The intermittent clamp technique used during CPB imposes an initial period of hypoxicischemic insult followed by reperfusion-reoxygenation damage, both likely to involve the early generation and release of oxidant species followed at a later stage by an inflammatory reaction [3,4]. In addition, extracorporeal circulation involves contact of blood with foreign surfaces which also leads to free radical generation and inflammation. Increased oxidant burden throughout CPB may overwhelm endogenous antioxidant defences and cause damage [3].



Figure 2. Time-course of changes in concentration of selenium and glutathione peroxidase (GPx) in arterial blood from adult patients subjected to cardiac surgery with cardiopulmonary bypass (CPB). Samples were obtained pre-operatively (T_0), after commencing the CPB (T_1), during ischemia (T_2 , T_3) and reperfusion (T_4 , T_5) and after surgery (T_6), as indicated in Methods. Data are mean \pm SD of 27 patients.

We have selected vitamins like ascorbate, α tocopherol and retinol, the trace element selenium and the selenoprotein GPx as relevants in defence against oxidant stress, although other key antioxidants have been recently studied in cardiac surgery [4]. Vitamin C appears as a particularly sensitive target of CPB-mediated oxidation in patients. Thus, plasma ascorbate levels were found markedly diminished in adult [23] and paediatric [4] patients after CPB. Others found that ascorbate concentrations, corrected for haemodilution, rose initially from those prior to surgery, remained elevated throughout CPB and returned to pre-operative levels by 24 h postsurgery [21]. We found an initial rise in plasma vitamin C followed by progressive decrease during reperfusion and a marked loss ($\sim 60\%$ decrease from baseline values) after surgery. The transient increase in vitamin C levels at the start of CPB may be due to its release from body stores as an early response to oxidant stress [24]. Alternatively, this augmentation may simply reflect the release to plasma of vitamin C from blood cells damaged by the extracorporeal circulation since blood cells have high intracellular levels of this antioxidant [25]. A direct correlation was found between pre-operative vitamin C concentrations and its decrease after surgery; although this relation has not been described by others, a similar finding was reported for total plasma antioxidant status [26].

 α -Tocopherol has been reported not to change significantly during CPB [4] or to decrease after reperfusion or after surgery [3]. By contrast, a transient increase of α -tocopherol at the end of CPB has been also reported [13]. We found a decrease of serum α -tocopherol throughout CPB and after surgery. Changes in retinol levels during CPB have been scarcely studied. No change of plasma retinol during CPB with decrease at 4 h post-surgery [3] or decrease during CPB [27] have been reported. We found a decrease in serum retinol



Figure 3. Correlations between the plasma levels of vitamin C, α -tocopherol and retinol at different stages of cardiopulmonary bypass surgery as shown in A, B and C. Points represent the absolute values at T_0 (\blacksquare), T_1 (\square), T_2 (\bigcirc), T_4 (\blacktriangle), T_5 (\triangle) and T_6 (\triangledown) as indicated in the legend of Figure 1 and Methods (n = 189 points in each graph).

which parallels that of α -tocopherol. Since we have not studied oxidant damage, it is not clear whether decreases of α -tocopherol and retinol indicate consumption of these lipid soluble vitamins or changes in their distribution pattern [28,29].

Plasma selenium was reported to decrease at 3 h after reperfusion but values were uncorrected for haemodilution [30]. Plasma GPx activity was found increased [3] or not changed [30,31] during CPB. We found no significant change for corrected values of plasma selenium and GPx during CPB and after surgery compared to pre-operative values.

The correlations between changes in endogenous antioxidant defences and clinical variables of CPB have been scarcely reported. In the present study, a significant correlation was found between maximal levels of PaO₂ during CPB and the extent of α -tocopherol and retinol loss after surgery. A similar tendency existed for vitamin C consumption but this correlation failed to reach significance. There is concern about whether excessive oxygenation in cardiac surgery induces oxygen-derived free radicals [32]. In fact, hyperoxic CPB in adults results in oxidative myocardial damage [33]. The basis for the relationship between maximal PaO₂ values and lipid soluble antioxidant vitamin levels found in this study remain uncertain since we did not investigate oxidant damage in this study. By contrast, we did not find any significant correlation between the amounts of loss in plasma antioxidant concentrations of vitamin C, α -tocopherol and retinol vs the duration of CPB, the duration of the ischemia and reperfusion periods and the degree of hypothermia. Although the duration of CPB has been shown to be an independent factor of CPB-associated post-operative complications [34], others have also failed in finding a correlation between total plasma antioxidant status and duration of CPB



Figure 4. Correlations between the lost in plasma levels of vitamin C (A), α -tocopherol (B) and retinol (C) expressed as a percentage of the initial values and the maximal values of PaO₂ (mmHg; \bullet), ischemic time (min; \triangle), reperfusion time (min; \bigtriangledown), CPB time (min; \square) and hypothermia (°C; \blacktriangle) during surgery for each patient (n=27 points for each clinical variable in each graph). (A) The loss of vitamin C did not correlate with PaO₂ (p=0.125, r=0.315), ischemic time (p=0.257, r=0.236), reperfusion time (p=0.896, r=0.030), CPB time (p=0.372, r=0.189) and hypothermia (p=0.553, r=0.128). (B) The loss of α -tocopherol correlates with PaO₂ (p=0.047, r=0.395) but not with ischemic time (p=0.199, r=0.255), reperfusion time (p=0.0704, r=0.078), CPB time (p=0.268, r=0.226) and hypothermia (p=0.064, r=0.382). (C) The loss of retinol correlates with PaO₂ (p=0.014, r=0.467) but not with ischemic time (p=0.348, r=0.188), reperfusion time (p=0.886 r=0.030), CPB time (p=0.537, r=0.127) and hypothermia (p=0.076, r=0.354).

but differentiation among endogenous antioxidants was not reported [26].

A limitation of our study is that we have not searched for direct evidences of oxidative injury by measuring biomarkers of lipid peroxidation [5]. Nevertheless, the progressive decrease of plasma non-enzymatic antioxidant levels from baseline is likely to reflect their consumption in relation to oxidant stress elicited by free radicals generated during the hypoxia and reoxygenation of myocardium. In fact, several clinical studies have examined the potential benefits of pre-operative antioxidant supplementation with vitamins C and E [10,35,36] as well as with other antioxidants [9,11] in the setting of CPB.

In summary, the present study shows that a significant consumption of plasma antioxidant vitamins occurs during and after CPB in adult patients, which correlates with arterial partial pressure of oxygen but not with other clinical variables measured in the present study. Optimization of surgical procedures, nutritional status and therapeutic interventions with antioxidant vitamins should therefore be considered as strategies to lower the rate of post-operative complications in patients undergoing cardiac surgery with CPB.

Acknowledgements

This work was supported by grants SAF2006-01002 (EJM) from CICYT (Ministry of Science and Technology, Spanish Government) and research aids from Research Foundation of University General Hospital Consortium (Valencia, Spain), CIBER CB06/06/ 0027 from Ministry of Health (Spain) and Regional Government (*Generalitat Valenciana*). The authors are indebted to the work of the surgical team of the University Hospital. Part of this work was presented in abstract form at the Meeting of the European Society of Parenteral and Enteral Nutrition (Madrid, Spain, 2000). The authors have no conflict of interest to declare.

References

- Cooley DA, Frazier OH. The past 50 years of cardiovascular surgery. Circulation 2000;102:IV87–IV93.
- [2] Murphy GJ, Angelini GD. Side effects of cardiopulmonary bypass: what is the reality? J Card Surg 2004;19:481–488.
- [3] Luyten CR, van Overveld FJ, De Backer LA, Sadowska AM, Rodrigus IE, De Hert SG, De Backer WA. Antioxidant defence during cardiopulmonary bypass surgery. Eur J Cardiothorac Surg 2005;27:611–616.
- [4] Christen S, Finckh B, Lykkesfeldt J, Gessler P, Frese-Schaper M, Nielsen P, Schmid ER, Schmitt B. Oxidative stress precedes peak systemic inflammatory response in pediatric patients undergoing cardiopulmonary bypass operation. Free Radic Biol Med 2005;38:1323–1332.

- [5] Cavalca V, Sisillo E, Veglia F, Tremoli E, Cighetti G, Salvi L, Sola A, Mussoni L, Biglioli P, Folco G, Sala A, Parolari A. Isoprostanes and oxidative stress in off-pump and on-pump coronary bypass surgery. Ann Thorac Surg 2006;81:562– 567.
- [6] Osaka M, Aoyagi K, Hirakawa A, Nakajima M, Jikuya T, Shigeta O, Sakakibara Y. Comparison of hydroxyl radical generation in patients undergoing coronary artery bypass grafting with and without cardiopulmonary bypass. Free Radic Res 2006;40:127–133.
- [7] Davies SW, Duffy JP, Wickens DG, Underwood SM, Hill A, Alladine MF, Feneck RO, Dormandy TL, Walesby RK. Time-course of free radical activity during coronary artery operations with cardiopulmonary bypass. J Thorac Cardiovasc Surg 1993;105:979–987.
- [8] Morcillo EJ, Estrela J, Cortijo J. Oxidative stress and pulmonary inflammation: pharmacological intervention with antioxidants. Pharmacol Res 1999;40:393-404.
- [9] Tossios P, Bloch W, Huebner A, Raji MR, Dodos F, Klass O, Suedkamp M, Kasper SM, Hellmich M, Mehlhorn U. N-acetylcysteine prevents reactive oxygen species-mediated myocardial stress in patients undergoing cardiac surgery: results of a randomized, double-blind, placebo-controlled clinical trial. J Thorac Cardiovasc Surg 2003;126:1513–1520.
- [10] Westhuyzen J, Cochrane AD, Tesar PJ, Mau T, Cross DB, Frenneaux MP, Khafagi FA, Fleming SJ. Effect of preoperative supplementation with alpha-tocopherol and ascorbic acid on myocardial injury in patients undergoing cardiac operations. J Thorac Cardiovasc Surg 1997;113:942–948.
- [11] Paparella D, Yau TM, Young E. Cardiopulmonary bypass induced inflammation: pathophysiology and treatment. An update. Eur J Cardiothorac Surg 2002;21:232–244.
- [12] Rastan AJ, Bittner HB, Gummert JF, Walther T, Schewick CV, Girdauskas E, Mohr FW. On-pump beating heart versus off-pump coronary artery bypass surgery—evidence of pump-induced myocardial injury. Eur J Cardiothorac Surg 2005;27:1057–1064.
- [13] Ulus AT, Aksoyek A, Ozkan M, Katircioglu SF, Vessby B, Basu S. Oxidative stress and changes in alpha- and gammatocopherol levels during coronary artery bypass grafting. Ann NY Acad Sci 2004;1031:352–356.
- [14] Ulus AT, Aksoyek A, Ozkan M, Katircioglu SF, Basu S. Cardiopulmonary bypass as a cause of free radical-induced oxidative stress and enhanced blood-borne isoprostanes in humans. Free Radic Biol Med 2003;34:911-917.
- [15] Gray A, McMillan DC, Wilson C, Williamson C, O'Reilly DS, Talwar D. The relationship between the acute changes in the systemic inflammatory response, lipid soluble antioxidant vitamins and lipid peroxidation following elective knee arthroplasty. Clin Nutr 2005;24:746–750.
- [16] Merino-Ramirez MA, Juan G, Ramon M, Cortijo J, Rubio E, Montero A, Morcillo EJ. Electrophysiologic evaluation of phrenic nerve and diaphragm function after coronary bypass surgery: prospective study of diabetes and other risk factors. J Thorac Cardiovasc Surg 2006;132:530–536.
- [17] Braubacher G, Vuilleumer JP. Vitamin C. In: Curtis HC, Rots M, editors. Clinical biochemistry. Vol 2. Berlin: Gruyter, 1974. p 989–997.
- [18] Driskell WJ, Neese JW, Bryant CC, Bashor MM. Measurement of vitamin A and vitamin E in human serum by highperformance liquid chromatography. J Chromatogr 1982;231: 439–444.
- [19] Alegria A, Barbera R, Clemente G, Farre R, Garcia MJ, Lagarda MJ. Selenium and glutathione peroxidase reference values in whole blood and plasma of a reference population living in Valencia, Spain. J Trace Elem Med Biol 1996;10: 223–228.
- [20] Belsten JL, Wright AJ. European Community—FLAIR common assay for whole-blood glutathione peroxidase

(GSH-Px); results of an inter-laboratory trial. Eur J Clin Nutr 1995;49:921–927.

- [21] Tangney CC, Hankins JS, Murtaugh MA, Piccione W Jr. Plasma vitamins E and C concentrations of adult patients during cardiopulmonary bypass. J Am Coll Nutr 1998;17: 162–170.
- [22] Gey KF, Brubacher GB, Stahelin HB. Plasma levels of antioxidant vitamins in relation to ischemic heart disease and cancer. Am J Clin Nutr 1987;45:1368–1377.
- [23] Ballmer PE, Reinhart WH, Jordan P, Buhler E, Moser UK, Gey KF. Depletion of plasma vitamin C but not of vitamin E in response to cardiac operations. J Thorac Cardiovasc Surg 1994;108:311–320.
- [24] Smith LJ, Houston M, Anderson J. Increased levels of glutathione in bronchoalveolar lavage fluid from patients with asthma. Am Rev Respir Dis 1993;147:1461–1464.
- [25] Evans RM, Currie L, Campbell A. The distribution of ascorbic acid between various cellular components of blood, in normal individuals, and its relation to the plasma concentration. Br J Nutr 1982;47:473–482.
- [26] McColl AJ, Keeble T, Hadjinikolaou L, Cohen A, Aitkenhead H, Glenville B, Richmond W. Plasma antioxidants: evidence for a protective role against reactive oxygen species following cardiac surgery. Ann Clin Biochem 1998;35(Pt 5):616–623.
- [27] Ochoa JJ, Vilchez MJ, Ibanez S, Huertas JR, Palacio MA, Munoz-Hoyos A. Oxidative stress is evident in erythrocytes as well as plasma in patients undergoing heart surgery involving cardiopulmonary bypass. Free Radic Res 2003;37: 11–17.
- [28] Newcomer ME, Ong DE. Plasma retinol binding protein: structure and function of the prototypic lipocalin. Biochim Biophys Acta 2000;1482:57–64.

- [29] Blatt DH, Leonard SW, Traber MG. Vitamin E kinetics and the function of tocopherol regulatory proteins. Nutrition 2001;17:799–805.
- [30] Lafont A, Marwick TH, Chisolm GM, Van Lente F, Vaska KJ, Whitlow PL. Decreased free radical scavengers with reperfusion after coronary angioplasty in patients with acute myocardial infarction. Am Heart J 1996;131:219–223.
- [31] Carlucci F, Tabucchi A, Biagioli B, Simeone F, Scolletta S, Rosi F, Marinello E. Cardiac surgery: myocardial energy balance, antioxidant status and endothelial function after ischemia-reperfusion. Biomed Pharmacother 2002;56:483– 491.
- [32] Ng CS, Wan S, Yim AP, Arifi AA. Pulmonary dysfunction after cardiac surgery. Chest 2002;121:1269–1277.
- [33] Ihnken K, Winkler A, Schlensak C, Sarai K, Neidhart G, Unkelbach U, Mulsch A, Sewell A. Normoxic cardiopulmonary bypass reduces oxidative myocardial damage and nitric oxide during cardiac operations in the adult. J Thorac Cardiovasc Surg 1998;116:327–334.
- [34] Wesselink RM, de Boer A, Morshuis WJ, Leusink JA. Cardiopulmonary-bypass time has important independent influence on mortality and morbidity. Eur J Cardiothorac Surg 1997;11:1141–1145.
- [35] Yau TM, Weisel RD, Mickle DA, Burton GW, Ingold KU, Ivanov J, Mohabeer MK, Tumiati L, Carson S. Vitamin E for coronary bypass operations. A prospective, double-blind, randomized trial. J Thorac Cardiovasc Surg 1994;108:302– 310.
- [36] Cavarocchi NC, England MD, O'Brien JF, Solis E, Russo P, Schaff HV, Orszulak TA, Pluth JR, Kaye MP. Superoxide generation during cardiopulmonary bypass: is there a role for vitamin E? J Surg Res 1986;40:519–527.